CE IVDR

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AZUL SALIVA DNA EXTRACTION KIT

DNA IN 40 MINS | GOOD YIELDS FOR USE IN PCR/SEQUENCING

PRODUCT BROCHURE

Cat No-DE110

ISO 13485 CERTIFIED

PRODUCT DESCRIPTION

AZUL Saliva DNA Extraction Kit is an easy and efficient system for the isolation of total DNA from Saliva Samples. This kit uses a silica-based spin column technology for isolating DNA from biological samples, thereby eliminating toxic phenol-chloroform extractions. The eluted DNA is suitable for all sensitive downstream applications such as qPCR and Next-Generation sequencing.

KIT COMPONENTS

Components	For 50 preps	For 25 preps
Lysis Buffer 1 (LB1)	35 mL	20 mL
Lysis Buffer 2 (LB2)	2 mL	1 mL
Proteinase K	3 mL	2 mL
Binding Buffer (BB)	25 mL	15 mL
Wash Buffer 1 (WB1)	30 mL	15 mL
Wash Buffer 2 (WB2)	25 mL	15 mL
Elution Buffer (EB)	4 mL	2 mL
Spin Column	50 (Pouch pack)	25 (Pouch pack)

SPECIFICATIONS

Format	Spin Column
Sample type	Saliva
Equipment	Microcentrifuge
Processing time	<40 mins
Processing volume	300µL - 1mL
Type	Total DNA
Sample storage	Eluted DNA should be stored at ≤ -20°C
Yield	2 - 25µg
Purity	A260/280 ≥ 1.8
Kit Storage	Room Temperature
Kit Validity	Viable for 1 year if stored at appropriate conditions

NOTE: *Check the Binding Buffer and Lysis Buffer for any salt precipitation before every use. Re-dissolve any precipitate by warming the solution to 37°C, then cool it back to room temperature before use.*

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DNA EXTRACTION PROTOCOL

1. In a microfuge tube, take around 300 µL - 1 mL of fresh Saliva samples or Saliva stored in mWRAPR Saliva Collection Tubes and add 300 µL - 700 µL of Lysis Buffer 1 (LB1).
2. To the tube, add 25 µL of Lysis Buffer 2 (LB2) and mix briefly by pipetting 2-3 times or vortex for 30 sec.
3. Add 50 µL of Proteinase K, mix well and incubate the tube at 56°C for 15 mins.
4. Centrifuge the tube at 13,000 rpm for 5 mins.
5. Carefully transfer the clear supernatant to a new 1.5 mL microfuge tube. Add 500 µL of Binding Buffer (BB) and mix the tube briefly by inverting it a few times.
6. Transfer 800 µL lysate to the spin column inserted in a collection tube and centrifuge at 12,000 rpm for 2 mins.
7. Discard the flow-through and place the purification column back into the collection tube. Repeat this step until the entire lysate has been transferred into the column and centrifuged.
8. Add 600 µL of Wash Buffer 1 (WB1) to the column and centrifuge at 12,000 rpm for 1 min.
9. Add 500 µL of Wash Buffer 2 (WB2) to the column and centrifuge at 12,000 rpm for 1 min to completely remove salts and impurities.
10. Transfer the purification column to a clean, sterile microfuge tube and add 30 µL -50 µL of Elution Buffer or DNase/RNase-free water to the center of the column.
11. Centrifuge the column at 12,000 rpm for 2 minutes.
12. Discard the purification column and store the eluted DNA at -20°C or -80°C until use.

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FLOW DIAGRAM OF DNA EXTRACTION PROTOCOL

Saliva Samples

300 µL - 1 mL of Saliva Samples

Add 300 µL - 700 µL Lysis Buffer 1

+

25 µL of Lysis Buffer 2

+

50 µL Proteinase K

Incubate at 56°C for 15 mins

Centrifuge at 13,000 rpm for 5 mins

Transfer clear supernatant to new tube

Add 500 µL Binding Buffer

Mix well

Transfer lysate to Spin column

Centrifuge at 12,000 rpm for 2 mins

Add 600 µL Wash Buffer 1

Centrifuge at 12,000 rpm for 1 min

Repeat this step with 500 µL Wash Buffer 2

Add 30 µL - 50 µL Elution Buffer

Centrifuge at 12,000 rpm for 2 mins

Eluted DNA

Store at -20°C or -80°C until use

TROUBLESHOOTING

PROBLEM	POSSIBLE CAUSES	SUGGESTED SOLUTIONS
Low DNA Yield	**Sample input:** Too much sample input or significantly less sample used.	Use less input material or increase the volume of the Lysis Buffer for better lysis. Use of ≥300 µL of sample is recommended for good DNA yield.
Low DNA Purity(A260/A280)	Improper sample handling results in ethanol or salt contamination	Make sure lysate and wash buffers have passed entirely through the matrix of the column. This may require centrifuging at a higher speed or longer time.
RNA Contamination	Too much sample used	**To remove RNA:** Perform in-column RNase I treatment or perform RNase I treatment post-purification (not provided in the kit), then re-purify the treated sample.
DNA Degradation	Use of samples not stored at appropriate conditions	**To prevent DNA degradation**: Immediately collect and lyse fresh saliva samples into a Lysis Buffer. Collect and store the fresh saliva samples in mWRAPR Saliva Solution to ensure stability & integrity of DNA and process later.

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ORDERING INFO

CATALOG NO	PRODUCT	PREP
DE101	AZUL Tissue DNA Extraction Kit	25/50 preps
DE102	AZUL Animal Cell Culture DNA Extraction Kit	25/50 preps
DE103	AZUL Bacterial DNA Extraction Kit	25/50 preps
DE104	AZUL Plasmid DNA Extraction Kit	25/50 preps
DE105	AZUL Plant DNA Extraction Kit	25/50 preps
DE106	AZUL Soil DNA Extraction Kit	25/50 preps
DE107	AZUL Blood DNA Extraction Kit	25/50 preps
DE108	AZUL Cell-free DNA Extraction Kit	25/50 preps
DE109	AZUL DNA Extraction Kit- Difficult samples	25/50 preps
DE110	AZUL Saliva DNA Extraction Kit	25/50 preps
DE111	AZUL Stool DNA Extraction Kit	25/50 preps
DE112	Quick AZUL Bacterial/Fungal DNA Extraction Kit	25/50 preps
DE113	AZUL Microbiome DNA Extraction Kit	25/50 preps
DE114	AZUL Gel DNA Extraction Kit	25/50 preps
DE115	AZUL FFPE DNA Extraction Kit	25/50 preps
DE116	AZUL Chloroplast DNA Extraction Kit	25/50 preps
DE117	AZUL Mitochondrial DNA Extraction Kit	25/50 preps
DE118	AZUL Pollen DNA Extraction Kit	25/50 preps
DE119	AZUL Fungal DNA Extraction Kit	25/50 preps
DE120	AZUL Sperm DNA Extraction Kit	25/50 preps
DE121	AZUL Skin DNA Extraction Kit	25/50 preps

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FEEDBACK

How did this kit perform?

Did AZUL Extraction Kit fulfill expectations required for your research?
Let us know by filling out the feedback form here

Or scan the QR code:

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